

Review of Active Solutions for Packaging Applications

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- § Turnover 272 M€ Personnel 2,470
- § Unique research and testing infrastructure
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VTT on the map



Active Packaging			
Releasers	Absorbers		
preservatives	oxygen		
flavours	moisture		
antioxidants	ethylene		
cabon dioxide	carbon dioxide		
ethylene	odour		

Intelligent Packaging
time temperature indicators
oxygen indicators
humidity indicators
carbondioxide indicators
spoilage indicators





Demand for **active and intelligent** packaging in the U.S. is forecast to expand 8.0 percent annually to **3.1 billion** € in 2017, well above total packaging demand growth.

Active packaging demand is expected to increase 5.7 percent annually to 2.0 billion € in 2017.

ACTIVE & INTELLIGEN (million	T PACKA dollars)	GING DI	EMAND		
				% Annu	al Growth
Item	2007	2012	2017	2007- 2012	2012- 2017
Active & Intelligent Packaging Demand	1308	2370	3480	12.6	8.0
Active Packaging:	1230	1635	2160	5.9	5.7
Gas Scavengers	384	610	870	9.7	7.4
Corrosion Control Packaging	374	405	485	1.6	3.7
Moisture Control Packaging	281	348	425	4.4	4.1
Other	191	272	380	7.3	6.9
Intelligent Packaging	78	735	1320	56.6	12.4

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By: marketsandmarkets.com Publishing Date: February 2011 Report Code: FB 1539



Active packaging	Active substances and mechanisms
Oxygen scavenger	Iron powder, ferrous carbonate, ascorbic acid, polyunsaturated polymers, ethylene methacrylate and cyclohexene methacrylate, benzyl acrylate, ascorbate, MXD-6 (aromatic polyamide) oxidation, enzymes (glucose oxidase, catalase, alcohol oxidase, superoxide dismutase), sulphites, unsaturated fatty acids (oleic, linoleic, linolenic), immobilization of yeasts in solid holders, photosensitive dyes, butylated hydroxytoluene (BHT), vitamins E and C, transition metal catalysts (platinum, cobalt, copper), rice extracts, catechol
Carbon dioxide scavenger	Calcium hydroxide, sodium hydroxide, potassium hydroxide, zeolites, active carbon
Carbon dioxide emitter	Ferrous carbonate, ascorbic acid, sodium bicarbonate
Ethylene scavenger	Potassium permanganate, palladium catalyst, zeolites, clays, Oya- stone, active carbon, silica gel, metallic oxides, tetrazine
Moisture scavenger	Silica gel, zeolites, propyleneglycol, calcium, barium and magnesium oxides, calcium sulphate, natural clays (such as montmorillonite), different salts, polyvinyl alcohol, molecular sieves, glycerol, polyacrylate salts, graft copolymers of starch
Ethanol emitter	Encapsulated ethanol, ethanol vapour generators

 Table 1. Active packaging technologies and mechanisms [6,8,9,10,12,14].

Vartiainen, Jari, Antimicrobial surface coatings in packaging applications, Surface Coatings. Rizzo, M. & Bruno, G. (eds). Nova Science Publishers. New York (2009), 45 - 91





Yildirim, S. Newsletter Transfer, 3-2010, Wädenswil, Switzerland

Oxygen coming from both sides











Produce affected by Ethylene Gas

ETHYLENE-PRODUCING COMMODITIES

Apples Apricots Avocados Cantaloupe Dried fruit Kiwifruit, ripe Mangoes Papayas

Peaches Pears Plums Tomatoes

ETHYLENE-SENSITIVE COMMODITIES

Bananas Bell peppers Broccoli Brussels sprouts Cabbage Carrots Cauliflower Cucumbers Eggplant Greens Green/snap beans Honeydew Kiwifruit, unripe Lettuce Okra Peas Plantains Spinach Squash Sweet potatoes Watermelon

Source: The Packer's 2005 Produce Availability & Morchandising Guide



Comparision of March 2010 {220 votes} & April 2014 {122 votes} Results) Gathered from SpecialChem4Polymers Community Results of the Survey

Antimicrobial packaging for food

§ Micro-organisms cause spoilage of foodstuffs

- §The use of traditional preservatives "inside" the food matrix is ineffective
- § Microbial growth occurs mainly <u>at the surface</u> of solid and semisolid foodstuffs
- § Antimicrobial effect can be directed at the surface of foodstuffs, for example, by using antimicrobial packaging materials which come into direct contact with foodstuffs



THE AMOUNT OF TRADITIONAL PRESERVATIVES CAN BE DECREASED





The global antimicrobial packaging market was valued at 6.51 billion € in 2015.

U.S antimicrobial packaging market volume by base material, 2012 - 2024 (Kilo Tons)







Type of substance	Example of substance
Organic acids	Propionic, sorbic, acetic, lactic, benzoic acid
Bacteriocins	Nisin, pediocins
Essential oils	Thymol, eugenol, cinnamic acid
Natural phenols	<i>p</i> -Cresol, hydroquinones, catechins
Enzymes	Lactoperoxidase, lysozyme, lactoferrin
Proteins	Conalbumin, cathepsin
Antioxidant phenolics	BHT (butylated hydroxytoluene), BHA
Isothiocyanates	Allylisothiocyanate, hypothiocyanite
Antibiotics	Natamycin
Antimicrobial peptides	Defensins, cecropins, attacins, magainins
Fungicides	Imazalil, benomyl
Chelatin agents	EDTA, pyrophosphate, citrates
Inorganics	Sulphites, sulphur dioxide
Metals	Silver, copper
Parabenes	Methyl, propylparaben

Table 3. Antimicrobial substances of potential use in food packaging [31].

Vartiainen, Jari, Antimicrobial surface coatings in packaging applications, Surface Coatings. Rizzo, M. & Bruno, G. (eds). Nova Science Publishers. New York (2009), 45 - 91





Yildirim, S. Newsletter Transfer, 3-2010, Wädenswil, Switzerland



Incorporation in package

Chemical immobilization in package



Ref. Han, 2000

Immobilization principle Natural antimicrobial Linker Spacer Linker Plasma induced anchor points BOPP

Y. Kourkoutas et al. / Food Microbiology 21 (2004) 377-397



Antimicrobial packaging



- Integration of antimicrobials agents into packaging materials
- To kill or to inhibit the pathogenic and spoilage microorganism



INHIBITION ZONE METHODS FOR QUALITATIVE ASSESMENT OF SURFACES WITH LEACHABLE ANTIMICROBIALS



- § The sample is placed on solid medium containing the test microorganism and incubated until growth is visible.
- § A clear zone surrounding the sample indicates antimicrobial diffusion from the sample and subsequent growth inhibition.







VIABLE COUNT METHODS

ASTM E2149 "Shake-flask test"

- § The samples are cut to test pieces.
- § The test pieces are incubated in a shake flask in bacterial suspension containing known number of test bacteria.
- § After incubation the number of viable bacteria in the suspension is measured by plating
- § The number of viable bacteria in the flask containing the antimicrobial sample is compared to that in the flask containing the non-treated reference.







Vartiainen, Jari, Antimicrobial surface coatings in packaging applications, Surface Coatings. Rizzo, M. & Bruno, G. (eds). Nova Science Publishers. New York (2009), 45 - 91

Antimicrobial substance	Packaging material	Target microbe	Food product	Reference
Sodium benzoate Potassium sorbate Nisin + EDTA	Caseinate films Chitosan films Keratin films	B. subtilis	-	[173]
Imazalil		A. niger		
Sorbic acid	Casein films Carnauba wax films	S. rouxii, A. niger	Papaya and apricot cubes	[174]
Tocopherol	Gelatin films	-	Margarine	
Citric acid	Pectinate, pectate, zein films	-	Nuts	
Mustard oil	Paper/ plastic bag	Penicillium spp., A. flavus, Endomyces fibuliger	Rye and wheat bread	[120]
Chitosan	Chitosan coating	S. aureus, L. monocytogenes, P. aeruginosa	Cheese	[175]
Chitosan	Chitosan coating	L. innocua, L. monocytogenes	Emmental cheese	[176]
Chitosan	Chitosan coating	Alternaria sp, Penicillium sp, Cladosporium sp.	Precooked pizza	[177]
Chitosan	Chitosan coatings and films	A. niger	-	[95]
Lysozyme	Chitosan films	S. faecalis, E. coli	-	[96]

Benzoic acid	Methylcellulose coatings	Zygosaccharomyces rouxii, Zygosaccharomyces mellis	Fruit preserves	[178]
Benzoic acid Sorbic acid	Methylcellulose coatings	P. notatum, Rhodotorula	-	[81]
	Methylcellulose/			
	chitosan coatings			
Nisin	Methylcellulose coatings	M. luteus	-	[179]
	Hydropropyl methylcellulose coatings			
Potassium sorbate Acetic acid	Hydropropyl methylcellulose films	Salmonella	Tomatoes	[180]
Nisin	Hydropropyl methylcellulose films	L. innocua, S. aureus	-	[60]
Pediocin	Cellulose casings Plastic barrier bags	L. monocytogenes	Turkey breast (fresh poultry), ham (processed meat), beef (fresh meat)	[181]
Potassium sorbate	Starch films	E. coli, salmonella	Chicken breast	[182]
Lactic acid	Alginate films	E. coli, salmonella, L. monocytogenes	Lean beef muscle	[183]

Glucose oxidase	Alginate films	-	Fish	[184]
p-Aminobenzoic acid	Whey protein	L. monocytogenes, E. coli,	Bologna slices	[185-187]
Potassium sorbate	based (WPI) films	salmonella	Sausage slices	
Acetic acid			Hot dogs	
Lactic acid		· •		
Nisin	Whey protein based (WPI) films	L. monocytogenes	-	[188]
Nisin + EDTA	Whey protein	B. thermosphacta, salmonella,	-	[188-189]
Lysozyme + EDTA	based (WPI) films	E. coli, L. monocytogenes,		
Propyl paraben		S. aureus		
Nisin	Soy protein isolate	L. plantarum, E. coli	-	[57]
Lysozyme	films			
+ EDTA	Corn zein films		-	
Nisin	Corn zein films	L. monocytogenes	Milk	[190]
Nisin	Corn zein films	L. monocytogenes	Chicken	[191]
Potassium sorbate	Corn zein films	S. aureus	Cheese	[46]
Sorbic acid	Corn zein films	L. monocytogenes	Cooked sweet corn	[192]
	Wheat gluten films			
Nisin (adsorbed)	LDPE films	L. monocytogenes	-	[68]
Acetic acid	Chitosan films	Enterobacteriaceae,	Bologna,	[193]
Propionic acid		Lactobacillus sakei, Serratia	cooked ham,	
Lauric acid		liqueficiens	pastrami	
Cinnamaldehyde				
Allyl-isothiocyanate	Labels	Moulds	Lettuce, turnip greens,	[194]
			fresh ground mince,	
			kiwifruit, strawberries,	
			rock cake, bread rolls	

Silver	Plastics		Cheese	[195]
Nisin Tocopherol	Vinyl acetate- ethylene copolymer binder (Elvace) on paper	M. flavus		[38]
Nisin Sorbic acid Potassium sorbate	PVDC films	L. monocytogenes	តា	[53]
Benzoic anhydride	LDPE films	Rhizopus stolonifer, Penicillium spp., A. toxicarius	Cheese	[72]
Benzoic acid Sorbic acid	Poly(ethylene-co- methacrylic acid) PEMA	Penicillium spp., A. niger		[73]
Grapefiuit seed extract	LDPE films	S. aureus, E. coli	Curled lettuce, Soybean sprouts	[196]
Potassium sorbate	LDPE films	Yeasts	-	[54]
Nisin Citric acid Lactic acid Malic acid Tartaric acid	Soy protein films	L. monocytogenes, E. coli, salmonella	-	[197]
Hexamethylenetetramine	LDPE films	Yeasts, lactic acid bacteria	Orange juice, Cooked ham	[44]
Benzoic anhydride	LDPE films	-	Cheese, Toasted bread	[32]
Nisin (adsorbed)	PVC films LLDPE films Nylon films	Salmonella	Broiler	[65]

Nisin,	Polyamide coatings	Total aerobic bacteria,	Fresh oyster,	[198]
Lacticin from	on LDPE films	coliform bacteria	Ground beef	
Lactococcus lactis				
Benzoic acid	Ionomer films	Penicillium spp., A. niger	-	[199]
Benzoyl chloride				[296]
Nisin (adsorbed)	Plastic film (70:30,	L. innocua, S. aureus	Sliced cheese and ham	[170, 200]
	PE:PA)			
	Conservation	L inner C manual		
	Greaseproof and	L. Innocua, S. aureus		
	moisture resistant			
Imazalil	Vinul agatata	Molde Devicillium	Hard and some hard	[122]
шпадаш	v myr acetate	words, Fencinum	chasses	[125]
Ciliare enterna	A diama la series la series de		Detetes	[107]
Silicon quaternary	Adnesive layer on	Heimininosportum solani	Potatoes	[127]
ammonium sait	(Cravera D 055)			
E () (D)	(Cryovac D-955)		T 1 1 11 //	[114]
Extracts (Kneum	LDPE films	-	Fresh curled lettuce	[114]
paimatum, Coptis			cucumber	
chinensis)				
Ag-substituted Zirconium		-		
Sorbic acid		E. coli, S. aureus, Leuconostoc mesenteroides		
Basil (linalool, methyl	LDPE films	E. coli, L. innocua	Cheddar cheese	[115,317,
chavicol)				324]
Nisin	Sorghum starch	Lactobacillus delbrueckii	-	[61]
	and flour films			

Chitosan Lactic acid/ sodium lactate	Chitosan coatings	Total aerobic psychrotrophic bacteria, lactic acid bacteria, yeast	Strawberry Lettuce	[204]
Nisin	Wheat gluten films	L. monocytogenes	Turkey bologna	[70]
Lysozyme Nisin Sodium benzoate	PVOH films	M. luteus, Alicyclobacillus acidoterrestris, S. cerevisiae	- 1	[37,150]
Chitosan	Yam starch films	Salmonella		[205]
Herbs (oregano, marjoram, thyme)	Plastic film		Hamburger meat patties, pizza	[206]
Essential oils (oregano, pimento)	Casein/ whey protein isolate films	E. coli, Pseudomones spp.	Beef muscle	[116]
Nisin	Hydroxypropylmet hylcellulose films	M. luteus	121	[207]
Potassium sorbate Sorbic acid	Celluse-based coating	Salmonella	Eggs	[208]
Benzoic acid Sorbic acid	Varnish on plastic films		Fresh meat and cheese	[209]
Allyl-isothiocyanate, imazalil	Cyclodextrin + PLA and PLA/PCL films	Penicillium commune, P. roqueforti, P. nalgiovense, P. verrucosum, P. caseifulvum, P. camemberti and K. maxianus	Cheese	[210]

Butylhydroxytoluene (BHT)	HDPE films	-	Fresh oat flaked cereal	[211]
Sorbic acid	Butter paper	Bacteria, yeasts, molds	Paneer from milk	[55]
Lysozyme	PVOH, nylon, cellulose triacetate (CTA) films	M. luteus	-	[148]
Propyl paraben	Styrene-acrylate copolymer coating on paper	S. cerevisiae	-	[33]
Potassium sorbate Citric acid	Starch-based coatings	Bacteria, molds and yeasts	Strawberry	[212]
Calcium sorbate	CMC/ paper	25.	Bread	[213-214]
Benomyl	Ionomer films	Fungi		[172]
Imazalil	LDPE films	3 5 .	Bell pepper	[124]
Imazalil	LDPE films	Molds	Cheese	[122]
Ethanol vapour	Silicon oxide sachet (Ethicap)	10 0	Bakery	[215]
Iodine potassium iodide, Sodium ortho- phenylphenate, Biphenyl, Diphenylamine, Dichloronitroaniline	Paper wraps	Trichothecium roseum, Glomerella cingulata, Penicillium expansum, Rhizopus stolonifer	Golden Delicious apples	[216]
Chitosan	BOPP films	E. coli, B. subtilis		[217]
Glucose oxidase	BOPP films	E. coli, B. subtilis	-	[218]
Lactic acid	Chitosan coated paper	B. subtilis	- 1	[219]

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$SO_2 + CO_2$	Cardboard box	Botrytis cinerea	Grape fruit	[272]
Sorbic anhydride	PE films	A. niger, Penicillium sp.	-	[276]
Lysozyme, nisin, grape fruit seed extract (GFSE), EDTA	Na-alginate, κ-carrageenan- based films	M. luteus, listeria, salmonella, E. coli, S. aureus	-	[277]
Nisin	PE/biopolymer (methylcellulose, hydroxypropyl methylcellulose, ĸ-carrageenan or chitacon)	M. luteus	-	[278]
Garlic oil, potassium sorbate, nisin	Chitosan films	S. aureus, L. monocytogenes, B. cereus	-	[281]
Nisin Chitosan	Vinyl acetate ethylene co- polymer coating on paperboard	Aerobic bacteria and yeasts	Pasteurized milk and orange juice	[284]
Lysozyme	Whey protein isolate (WPI) films and coatings	L. monocytogenes	Cold-smoked salmon	[286]
Nisin + grape seed extract (GSE) green tea extract (GTE)	Soy protein films and coatings	L. monocytogenes	Turkey frankfurters	[287]
Bacteriocin from Lactobasillus curvatus (adsorbed)	PE films	L. monocytogenes	Pork steak, ground beef	[288]
Trans-cinnamaldehyde	Polyamide coating on LDPE films	L. innocua	Fresh-cut romaine lettuce	[292]

Chlorine dioxide Allyl isothiocyanate	Sachets in multilayer barrier tray	L. monocytogenes, salmonella	Chicken breast	[303]
2E-hexenal	Cyclodextrin + PLA	Fungi	Blueberries	[306]
Ion-exchanged zeolite	PE film	E. coli, salmonella, S. typhimurium	Iceberg lettuce	[307]
Lysozyme	Chitosan films	L. monocytogenes, E. coli, P. fluorescens, molds, yeasts	Mozzarella cheese	[311]
Antimicrobial peptide dermaseptin K ₄ K ₂₀ -S4	Starch coating on PE films	Molds, aerobic bacteria	Fresh cucumber	[314]
Natamycin	Celluse-based film	Penicillium roqueforti	Gorgonzola cheese	[315]
Enterocins	Alginate, zein and PVA films	L. monocytogenes	Sliced cooked ham	[320,321]
Nisin, sodium lactate, sodium diacetate, potassium sorbate, sodium benzoate	Chitosan coatings on Surlyn film	L. monocytogenes	Ham steaks	[325]
Allyl isothiocyanate	Labels in packaging	P. commune, P. roqueforti, A. flavus	Cheese	[327]
Sorbic acid	Cellulose polymer film	Mesophilic and psychotropic bacteria, Staphylococcus spp.	Pastry dough	[330]
2-nonanone	PP/EVOH cups	Botrytis cinerea	Strawberries	[331]
Cinnamaldehyde – enriched cinnamon essential oil	Paraffin coated paper	Fungi	Strawberries	[333]
Oregano	PE film	Yersinia enterocolitica	Beef meat	[335]
Nisin	PP/PA/PP interleavers	Salmonella	Cooked ham	[345]
Natural antimicrobial and antioxidant compounds



Chemo-enzymatic functionalisation of **VIT** lignin-containing fibre materials

PRINCIPLE OF THE METHOD

- § Based on laccase-catalysed oxidation of phenolic -OH to phenoxy radicals
- § Activation + Bonding = Functionalisation
- § Functional chemicals selected based on target modification

FUNCTIONAL PROPERTIES ADDED TO FIBRES:

- § Charge
- § Hydrofobicity
- § Conductivity
- § Traceability
- § Antimicrobial activity





Adjusting the level of hydrophobicity

Hydrophobicity: isoeugenol < octyl gallate < dodecyl gallate







Adjusting the level of hydrophobicity







Laccase-mediated grafting of phenolic antimicrobials



Elegir et al. (2008) Enzyme and Microbial Technology

Antimicrobial papers



Releases the active ingredient Benzalkonium chloride when in contact with wet hands





Low-cost, compostable and infused only with organic spices, FreshPaper keeps produce fresh for 2-4x longer.



Prevention of germ growth
Inhibition of mould (Candida albicans)
Hygiene maintenance and significant bacteria reduction (E.coli, S.aureus)
Odour control
Increase in shelf life and product freshness

Antimicrobial plasters





Antimicrobial bottles



Both containers and labels have proven efficacy across a broad spectrum of bacteria including MRSA and E. Coli. Both are based on ionic silver, a natural compound that kills over 99.9% of bacteria and provides food safe contact. Bottles and jars are available in SAN, PP, PET and HDPE.

Antimicrobial sol-gel precursors







Octadecylammoniumtrimethoxysilane, ODAMO Mercaptopropyltrimethoxysilane MPTMO

Antimicrobial activity of surface treated with solgels; ODAMO (SO154) and MPTMO (SO151)



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 Table 1. Antimicrobial activity^a of LDPE plates expressed as a mean diameter (mm) of inhibition zone.

Type of Plate (Thickness 2 mm)	Bacillus subtilis (1 day @ 30°C)	Aspergillus niger (2 days @ 30°C)
LDPE	-	-
LDPE + 0.05% imazalil	-	15
LDPE + 1% EDTA	-	-
LDPE + 0.025% imazalil + 0.5% EDTA	-	13
LDPE + 0.25% imazalil	-	40
LDPE + 5% EDTA	32	-
LDPE + 0.125% imazalil + 2.5% EDTA	35	19

^aMeans (n = 3).



Figure 5. Structure of (a) 0.02% imazalil; (b) 0.1% imazalil; (c) 0.2% imazalil; (d) 0.5% EDTA; (e) 2.5% EDTA and (f) 5% EDTA containing LDPE films (Olympus BX 50 microscope equipped with PCO CCD IMAGING SenseCam 12 BIT COOLEDIMAGING).

Vartiainen et al. (2003) Journal of Plastic Film and Sheeting. Vol. 19 (2003) No: 4, 249 - 262



Table I. Antimicrobial activity^a of plastic plaques expressed as a mean diameter (mm) of inhibition zone

Type of plaque (thickness 2mm)	Bacillus subtilis	Aspergillus niger
LDPE LDPE + 15% sodium benzoate LDPE + 15% potassium sorbate LDPE + 15% sodium nitrite LDPE + 15% sodium lactate	- 26 16 44	- - 42 -
MA MA + 15% sodium benzoate MA + 15% potassium sorbate MA + 15% sodium nitrite MA + 15% sodium lactate	- 30 15 44	- - 25 -
PS PS + 15% sodium benzoate PS + 15% potassium sorbate PS + 15% sodium nitrite PS + 15% sodium lactate	47 23 51	- - 32 -
PET PET + 15% sodium benzoate PET + 15% potassium sorbate PET + 15% sodium nitrite PET + 15% sodium lactate	- 48 31 65 -	- - 40 -

^a Means (n = 3).



Figure 5. Structure of (a) 2.5% sodium benzoate, (b) 7.5% sodium benzoate, (c) 15% sodium benzoate, (d) 2.5% sodium nitrite, (e) 7.5% sodium nitrite and (f) 15% sodium nitrite containing LDPE films (Olympus BX 50 microscope equipped with PCO CCD IMAGING SenseCam 12 BIT COOLED IMAGING).



Roll-to-roll treatments in coating pilot-line



Plasma activation



 § Plasma is a (partially) ionised gas in which ions and electrons are present as well as radicals and molecules in an excited state
 § Plasma contains both physical as well as chemical very reactive species.







Plasma activation





Surface energy (polar component mN/m)





Wet-coating of plasma-activated **v**

§Biobased polymers modified with antimicrobial compounds and enzymes







20/06/2017





Figure 4. Surface topography of (a) untreated, (b) N_2 -plasma + NH_3 treated and (c) chitosan coated, N_2 -plasma + NH_3 pretreated BOPP films and (d) cross-section of chitosan coated sample.

Table 3. Gas transmissions of BOPP films expressed as cm³/(m²·24 h).

Film	Oxygen	Carbon dioxide	Ethylene
BOPP (without plasma)	1500	4000	430
BOPP (N ₂ -plasma + NH ₃)	1500	4600	390
Chitosan coated BOPP (N2-plasma + NH3)a	27	790	53

Means (n=2)



Figure 5. Effect of plasma activated BOPP film with or without surface immobilized chitosan (0.1% glutaraldehyde as linking agent) on survival of *E. coli* and *B. subtilis* (24 h in peptone saline at 30 °C). Means (n=3).



Figure 1 (A) Uncoated and (B) chitosan/6.4% propionic acid coated boards (taken with an Olympus BH-2 microscope equipped with an Olympus DP12 microscope digital camera system).

Vartiainen et al. (2004) Journal of Applied Polymer Science. Vol. 94 No: 3, 986 - 993



Vartiainen et al. (2007) 10th Pacific Polymer Conference (PPC 10). Kobe, Japan, 4 - 7 Dec. 2007

Figure 3. Proposed reactions in tyrosinase-catalysed grafting of gallates to chitosan.

(1)

(2)

-chitosan

-chitosan

chitosan

tyrosinas + O₂



Figure 4. SEM cross-section picture of chitosan-coated plasma-activated BOPP film.

Vartiainen et al. (2008) Packaging Technology and Science. Vol. 21 No: 6, 317 - 328



Development of antimicrobial packaging materials with immobilized glucose oxidase and lysozyme

Research Article

Kristýna Hanušová1*, Lukáš Vápenka1, Jaroslav Dobiáš¹, Linda Mišková² Storage time Activity of immobilized GOX (%) (day) Polyamide film lonomer film Polyamide film (A) -CO 100 100 0 NH (B) lonomer film 97.5 ± 1.1 14 98.3 ± 1.4 Ç0 21 97.1 ± 1.7 952 ± 19 Enzyme HC-CO-N 28 93.6 ± 1.4 80.9 ± 1.8 (CH_), 41 92.1 ± 2.3 77.6 ± 2.0 HC-CO (CH_), 56 73.8 ± 2.2 60.4 ± 1.3 CO -co-ċh Enzyme 193 17.5 ± 4.2 15.0 ± 2.5 CO 5.4 ± 1.6 3.1 ± 1.4 245 Enzyme

The scheme of linkage between enzyme and polyamide film (A) and ionomer film (B) with indication of hypothetical cleavage (dashed curve) of enzyme from polymer.



Figure 5. The growth inhibition of indicator bacteria: Escherichia coli (A) and Pseudomonas fluorescens (B) by ionomer film with GOX; Lactobacillus helveticus (C) and Listeria innocua (D) by polyamide film with GOX; Listeria ivanovii (E) by polyamide film with GOX during cold storage at 4°C; Bacillus subtilis (F) by cellulose film releasing LYS.









Glucose oxidase immobilized onto N₂-plasma + NH₃ activated PP







Figure 3. Schematic diagram of immobilization of enzyme (glucose oxidase) onto (A) carboxyl-activated and (B) amino-activated food packaging film (BOPP).





Figure 9. Antimicrobial activity of BOPP films immobilized with glucose oxidase. Immobilization was carried out in phosphate buffer, pH 7.2, containing 2.5% of glutaraldehyde for 10s at 25°C, 55°C, 65°C and 75°C, or for 1 day at 4°C. Films were incubated for 24h at 30°C. Means (n = 2).



Vartiainen et al. (2005) Packaging Technology and Science. Vol. 18 (2005) No: 5, 243 - 251 Chicken breast fillets packed into plasma-activated film immobilized with two natural antimicrobial peptides





Gelatin coating with two natural antimicrobial peptides

- Speed 5 m/min
- Corona treatment prior to gelatin coating
- Coating was applied onto PE surface
- Flexo-coating unit was tempered (50°C)







Gelatin coating with two natural antimicrobial peptides

- Drying with 5 air dryers at 80°C
- Totally 738 m (top film; width 355 mm) and 461 m (down film; width 360 mm) were coated with gelatin
- Approx. 1 g/m²

Antimicrobial vacuum packaging for fish







Vacuum-packed fresh salmon fillet

- Pouches of the dimension 25 x 15 cm were prepared.
- Fresh salmon fillets, with a weight of approximately 200 g each, were packaged individually and sealed under vacuum.
- Microbiological, chemical and sensory analyses were carried out after storing at 2°C for 8 days.















Antimicrobial activity



Some examples of VTT research results

Antimicrobial treatments for sausage casings

• Activity against Listeria



- Antimicrobial printing inks for bread packaging
 - Activity against bread spoiling moulds





Immobilization procedure using atmospheric plasma


Antimicrobial printing



Target: To develop ink with antimicrobial properties applicable with conventional printing technique (flexo or rotogravure) to food packaging material.



Inner surface of the packaging material is covered with an antimicrobial ink layer coming in contact with packed foodstuffs.









Antimicrobial printing: advantages

§Thin active layers

§total amounts of antimicrobial substances as low as possible

§Traditional printing method

§flexo or rotogravure

§No high temperatures

§higher antimicrobial activity

§Easy to apply

§whenever, wherever...



Functionalization of paper by adding inorganic TiO₂ and ZnO nanoparticles

- TiO₂ and ZnO have photo-oxidative and antimicrobial activities
 - Inhibition to grow (bacteriostatic) or killing (bactericidal) of patogenic bacteria: gram-, gram+ and bacteria spores.
- TiO₂ 40 nm nanoparticles and ZnO 45 nm nanoparticles







Nano-composite	Initial mixing ratio of the preparations (as dry weight)	TiO ₂ content (% dry weight)	ZnO content (% dry weight)
CNF-TIO ₂	1:1	24	-
CNF-ZnO	10 : 1	-	7
CNF-ZnO&TiO ₂	10 : 1 : 10	42	2

Nano-composites preparation:

- 15 min mixing CNF suspension (2.5% in water) with TiO₂ (6% in water) and/or ZnO (1% in ethylene glycol)
- 3 washings with water followed by centrifugation to eliminate not-adsorbed inorganic fillers

Antimicrobial paper



Antibacterial activity under conditions of light exposition. Bacteria grow on untreated reference: $10^6 - 10^7$ (6-7 log).

Paper Samples	ppm ZnO	% TiO₂	S. aureus (Gram +)		K. pneumoniae (Gram -)	
			Bacteriostatic activity (log reduction)	Bactericidal activity (log reduction)	Bacteriostatic activity (log reduction)	Bactericidal activity (log reduction)
Untreated paper (control)	0	0	0	0	0	0
Paper CNF-TiO ₂ (0.4 g/m ²)	0	0.034	0	0	0	0
Paper CNF-TiO ₂ (0.9 g/m ²)	0	0.167	1.3	0	0	0
Paper CNF-TiO ₂ (1.8 g/m ²)	0	0.291	Total bacteriostatic	1.0	1.8	0
Paper CNF-ZnO&TiO ₂ (0.6 g/m²)	38.4	0.115	0.7	0	0	0
Paper CNF-ZnO&TiO ₂ (1.3 g/m ²)	104.6	0.307	Total bacteriostatic	0.1	0.2	0
Paper CNF-ZnO (0.8 g/m²)	180	0	Total bacteriostatic	0.3	2.1	0

Significant bacteriostatic activity was obtained (inhibition to bacteria proliferation) with TiO₂ contents around 0.3%

• Bactericidal activity (bacteria killing-reduction of initial inoculated cells) was not detected.

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Active paper for the photo-oxidation of volatile substances in gaseous phase

Kinetic of NOx photodegradation in gaseous phase by CNF-TiO₂ foam coated paper



Around 70% NOx oxidation within 100 minutes

ØCNF-TiO₂ has significant activity for the oxidation of NO and NOx

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ALD; Repeated Self-Limiting Gas-Solid Reactions







Antimicrobial activity

Thin coatings	Activity
25 nm Al ₂ O ₃ (water) 70 °C	3.5
25 nm ZnO (water) 70 °C	5.9
50 nm ZnO (water) 70 °C	5.8
75 nm ZnO (water) 70 °C	5.8
50 nm ZnO + 5nm Al ₂ O ₃ 70 °C	5.9
50 nm ZnO + 15nm Al ₂ O ₃ 70 °C	4.6
25 nm ZnO (water) 100 °C	2.3
50 nm ZnO (ozone) 70 °C	5.8
50 nm ZnO (ozone) 100 °C	5.8

§ Determined with JIS Z 2801,

- § Antimicrobial if activity > 2,
- § All samples antimicrobial against Escherichia coli,
- § Direct contact rather than edge diffusion more efficient,
- § Nanolaminate consisting of 50 nm ZnO covered with a layer of Al₂O₃ retained most of its antimicrobial activity compared to plain ZnO.



Antimicrobial activity



- § 50 nm of ZnO deposited onto BOPLA with water at 70 °C indicated lateral diffusion of an antibacterial agent from a relative thick film
- § Growth of Aspergillus niger and Bacillus subtilis inhibited
- § EN 1104 (Hemmhof test) test commonly applied to evaluate materials for food contact



